

Thiolate S-Oxygenation Controls Nitric Oxide (NO) Photolability of a Synthetic Iron Nitrile Hydratase (Fe-NHase) Model Derived from Mixed Carboxamide/Thiolate Ligand

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Nitrile hydratase (NHase) is an iron- or cobalt-containing enzyme that catalyzes the hydrolysis of nitriles ($R-C\equiv N$) to the corresponding amides [$R-(C=O)-NH_2$].¹ It contains a non-heme Fe(III) or non-corrinoid Co(III) center bound to a unique array of ligands at the active site. The crystal structure of iron NHase (Fe-NHase) revealed an Fe(III) center in an octahedral geometry. In the equatorial plane, it is coordinated to two unusual nitrogen donors derived from the peptide backbone (the deprotonated carboxamido N atoms from Cys112 and Ser113) and two cysteinato S donors that are post-translationally oxygenated to sulfenate (SO; Cys114) and sulfinato (SO₂; Cys112) moieties, while the axial site is occupied by a third cysteine S donor (unmodified Cys110). The solvent-exposed sixth site is occupied by H₂O (or OH⁻) in the catalytically active enzyme. Interestingly, Fe-NHase can also be isolated in an inactive “dark form” when purified under strict low-light conditions. The 1.7 Å structure of dark-form Fe-NHase (Fe-NHase_{dark}) revealed a single molecule of nitric oxide (NO) bound to the active-site iron,² thus inhibiting enzymatic activity. Upon exposure to light, NO is photoreleased and catalytic activity is restored, suggesting that Fe-NHase is photoregulated by NO.

Synthetic work by several groups has generated small-molecule models of Fe-NHase_{dark}. For example, Kovacs and co-workers³ have employed a Schiff base ligand containing imine N and thiolato S donors to generate the six-coordinate nitrosyl [(S₂Me₂N₃^{PtPr})-Fe(NO)]⁺, which emulates the NO-bound active site of Fe-NHase. Another model complex, namely, [(btmp-TASN)Fe(NO)]⁺, reported by Grapperhaus et al.,⁴ also exhibits an Fe-bound NO moiety, but *neither complex exhibits NO photolability*. Artaud and co-workers⁵ have incorporated carboxamido N and thiolato S donors in the nitrosyl [(N₂S₂)Fe(NO)]⁻. However, the complex is only five-coordinate and also does not exhibit any NO photolability. No model complex to date emulates the six-coordinate {Fe-NO}⁶ active site with bound carboxamide and thiolate groups that is observed in Fe-NHase_{dark}. As a consequence, the chemical factors that lead to NO photolability remain unclear. In this work, we for the first time report a model of dark-form Fe-NHase with carboxamido N and thiolato S coordination. In addition, photorelease of NO from the S-oxygenated model complex is reported.

Reaction of deprotonated Cl₂PhPepSH₄ (NaH/DMF) with NEt₄[FeCl₄] afforded a red-brown solution of the Fe(III) precursor [(Cl₂PhPepS)Fe(Cl)]²⁻.⁶ Addition of 3 equiv of DMAP at room temperature immediately generated a dark-green color, indicating substitution of Cl⁻ by DMAP. Storage of this complex in THF/Et₂O at -20 °C for several weeks afforded dichroic green/red needles of NEt₄[(Cl₂PhPepS)Fe(DMAP)] (1). The IR spectrum of 1 (KBr disk; Figure S1 in the Supporting Information) exhibits a ν_{CO} peak at 1590 cm⁻¹, which is typical for metal-bound carboxamide. The X-ray structure of 1 (Figure 1) reveals a five-coordinate structure wherein the deprotonated N₂S₂ ligand is bound to the Fe(III) center in the equatorial plane and the axial site is occupied by DMAP (ν_{CN} = 1614 cm⁻¹). Complex 1 exhibits an EPR signal with features at g = 4.39 and 1.98 (MeCN/toluene glass, 125 K;

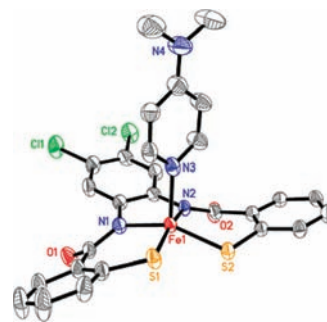


Figure 1. Thermal ellipsoid plot (50% probability level) of [(Cl₂PhPepS)Fe(DMAP)]⁻ (the anion of 1). Selected bond distances (Å): Fe–N1, 1.949(3); Fe–N2, 1.965(3); Fe–S1, 2.1994(11); Fe–S2, 2.2065(11); Fe–N3, 2.108(4).

Figure S2) and μ_{eff} = 3.74 μ_B at 298 K, both of which are typical values for square-pyramidal S = 3/2 Fe(III) systems.

Complex 1 displays a strong affinity for NO in solution. When a green (λ_{max} = 650 nm) solution of 1 in MeCN at -40 °C was treated with 1 equiv of NO gas, an immediate color change to pale-red was observed. Storage of the solution (1:3 MeCN/Et₂O) at -40 °C for several days afforded a red microcrystalline material. The IR spectrum of the resulting nitrosyl exhibits a strong ν_{NO} stretch at 1849 cm⁻¹ (KBr disk; Figure S3) characteristic of an {Fe-NO}⁶ system. The IR spectrum also exhibits a feature at 1624 cm⁻¹ due to bound DMAP. The X-ray structure (Figure 2) confirms that the resulting nitrosyl is indeed the monomeric species NEt₄[(Cl₂PhPepS)Fe(NO)(DMAP)] (2). The iron center of 2 is coordinated to the two carboxamido N and two thiolato S donors in the equatorial plane. The extreme distortion of the planar ligand frame results in asymmetric Fe–N bond distances [Fe–N1 = 1.944(9) Å; Fe–N2 = 2.007(11) Å], while the Fe–S bond distances are quite similar [Fe–S1 = 2.273(5) Å; Fe–S2 = 2.279(3) Å]. The nearly linear Fe–N–O unit [173.2(8)°] resembles that observed in the native dark-form enzyme (av 165°), and the corresponding bond distances [Fe–N3 = 1.612(10) Å; N3–O3 = 1.167(11) Å] are typical of an

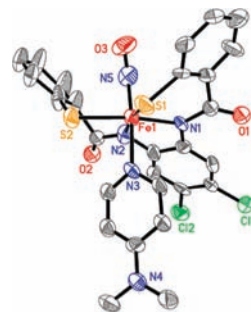


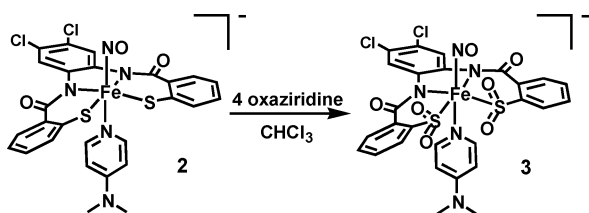
Figure 2. Thermal ellipsoid plot (50% probability level) of [(Cl₂PhPepS)Fe(NO)(DMAP)]⁻ (the anion of 2). Selected bond distances (Å) and bond angles (deg): Fe–N1, 1.944(9); Fe–N2, 2.007(11); Fe–S1, 2.273(5); Fe–S2, 2.279(3); Fe–N3, 1.612(10); Fe–N5, 1.612(10); N5–O3, 1.167(11); Fe–N5–O3, 173.2(8).

{Fe–NO}⁶ core. The bound DMAP is trans to NO and exhibits a shorter Fe–N bond distance [Fe–N3 = 2.065(8) Å] than that observed in **1** [Fe–N3 = 2.108(4) Å].

The nitrosyl **2** exhibited interesting NO reactivity in solution *under dark conditions*. When red crystals of **2** were redissolved in MeCN (at room temperature or –40 °C), the solution immediately turned green, indicating formation of **1**. This was confirmed by the electronic absorption, EPR, and IR spectra of the green product. Repeated NO addition followed by evacuation at –40 °C cleanly cycled between **1** (λ_{\max} = 590 nm) and **2** (λ_{\max} = 610, 950 nm; Figure S4). This shows that the bound NO in **2** is *labile* and readily dissociates in MeCN and other coordinating solvents such as THF or DMF. In contrast, **2** can be dissolved in noncoordinating solvents (e.g., CHCl₃ or CH₂Cl₂), where the red solutions are stable even at room temperature.

Next, we tested the photoactivity of **2**. When red solutions of **2** in noncoordinating solvents were exposed to visible or UV light, no change in the electronic absorption spectrum was observed. This clearly indicates that *NO is not photolabile* from **2**, despite its structural similarity (octahedral, carboxamido N, thiolato S) to the active site of Fe–NHase_{dark}. Also, the red solution of **2** (λ_{\max} = 610, 950 nm) bore little resemblance to that of the orange enzyme (Fe–NHase_{dark}; λ_{\max} ≈ 400 nm). The lack of NO photolability suggested that further modification of **2** was required to accurately model the photoactive {Fe–NO}⁶ core of Fe–NHase_{dark}.

It has been proposed by us⁷ and others^{2,4} that S-oxygenation of cysteine thiolates may regulate the photolability of NO from Fe–NHases. In the present work, we explored this possibility by attempting to oxygenate the thiolato S donors of **2**. Treatment of **2** with a variety of standard oxygenating agents (O₂, H₂O₂, and dimethyldioxirane) at –40 °C led only to decomposition of the reaction mixture (no isolable species). However, we found that treatment of **2** with 4 equiv of (1*S*)-(+)-(10-camphorsulfonyl)oxaziridine in CHCl₃ *in the dark* afforded a pale-orange solution with λ_{\max} = 440 nm (compare to λ_{\max} ≈ 400 nm for Fe–NHase_{dark}; see Figure 3). This species was stable only for limited periods of time in MeCN, CH₂Cl₂, or CHCl₃ at temperatures as low as –40 °C.



Unlike **2**, the oxygenated species *exhibits no lability of NO* under vacuum. The resulting IR spectrum (Figure S5) displays a slightly shifted ν_{NO} value of 1854 cm^{–1} (enzyme ν_{NO} = 1853 cm^{–1}) as well as features characteristic of S-oxygenation (ν_{SO} = 1078, 1046, 1007 cm^{–1}), similar to those observed in Fe– and Co–sulfonates such as [(Cl₂N₂{SO₂})₂Fe(CN)₂]^{3–}, [(PyP{SO₂})₂Fe(CN)₂]^{2–}, and [(LN₂{SO₂})₂Co(*t*BuNC)₂][–] (all of which exhibit ν_{SO} = 1000–1200 cm^{–1}).^{8–10} Also, the ν_{CO} stretch at 1599 cm^{–1} and ν_{CN} at 1625 cm^{–1} indicate that the deprotonated Cl₂PhPepS^{4–} ligand and DMAP remain tightly bound to the Fe–NO unit. On the basis of the spectroscopic (Figure S5) and mass spectral (m/z = 721.1) data, we assign this species as [(Cl₂PhPep{SO₂})₂Fe(NO)(DMAP)][–] (**3**).¹¹

Most interestingly, exposure of a fresh solution of **3** in MeCN or CHCl₃ at –40 °C to low-intensity (10 mW) visible light immediately generated a pale-green solution with a new peak at 650 nm, similar to the behavior of the light-activated enzyme. Quantitative photochemistry in the visible region (λ_{irr} = 450 nm) afforded a quantum yield value of ϕ_{450} = 0.55, which is close to the quantum efficiency observed for

Fe–NHase_{dark} (ϕ_{355} = 0.48).¹² The fact that the color change was due to photorelease of NO from **3** was confirmed by measurement with a NO-sensitive electrode (Figure S6). The IR spectrum of irradiated **3** also confirmed the loss of NO from the complex. EPR measurements revealed the formation of low-spin [(Cl₂PhPep{SO₂})₂Fe(DMAP)₂][–] (g = 2.23, 2.03, and 2.02 and m/z = 823.1; Figure S7) as the photoproduct in the irradiated solution (generated at –40 °C in presence of 2 equiv of DMAP).

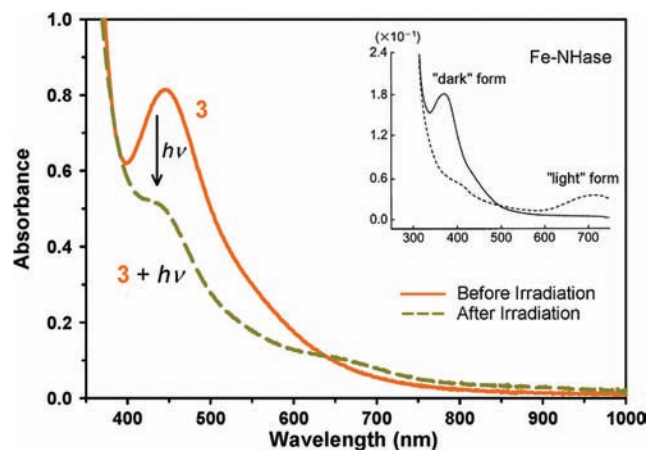


Figure 3. Electronic absorption spectrum of S-oxygenated {Fe–NO}⁶ nitrosyl **3** (MeCN, –40 °C), indicating its similarity to the spectrum of native Fe–NHase_{dark} (shown in the inset). Solid lines are spectra taken under dark conditions, while dashed lines represent spectra after exposure to light.

In summary, we have isolated a new monomeric iron nitrosyl derived from a mixed carboxamide/thiolate ligand that bears a unique resemblance to the “dark-form” Fe–NHase bound to NO. While NO is not photolabile from unmodified **2**, NO is photolabile from the S-oxygenated species **3**, suggesting an important role for S-oxygenation in controlling NO photorelease from Fe–NHase. Structural characterization of **3** is in progress.

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Supporting Information Available: IR spectra of **1** (Figure S1), **2** (Figure S3), and **3** (Figure S5); X-band EPR spectra of **1** (Figure S2) and the photoproduct of **3** (Figure S7); electronic absorption spectra showing the reversible formation of **1** and **2** (Figure S4); NO amperogram of **3** (Figure S6); and X-ray crystallographic data (CIF) for **1**·THF·Et₂O and **2**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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